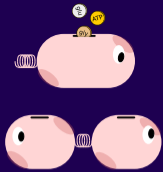


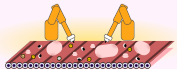
# Economic Principles in Cell Physiology

Paris, July 6, 2022

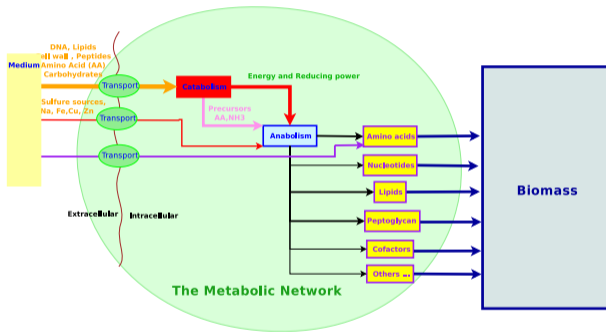


## Resource allocation models

**Anne Goelzer & Wolfram Liebermeister**



# The cell as considered by FBA



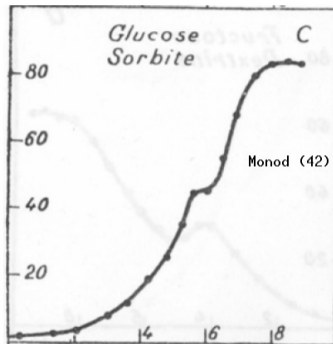
maximize  $c^T \nu$  (growth rate)  
 $\nu \in \mathbb{R}^m$   
subject to

$$\Omega \nu = 0$$

$$\alpha_i \leq \nu_i \leq \beta_i$$

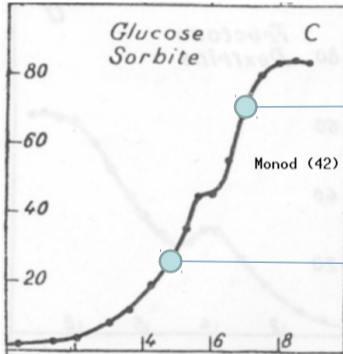
where  $\Omega$  is the stoichiometry matrix and  $c$  is the objective function (as growth rate maximisation).

# An effective modeling framework ...



Can FBA predict some well-established phenomena (as the catabolic repression)?

# Monod's experiments is not recovered by FBA



Predicted use of carbon source by other constraint-based modeling methods (FBA)

Sorbitol + Glucose

Sorbitol + Glucose

A relevant modeling framework, but unable to predict some well-established biological phenomena in an autonomous way.

# Advantages and drawbacks of Flux Balance Analysis

## Advantages

- ▶ no parameters;
- ▶ include the whole metabolic network through mass conservation:  $\Omega \cdot \nu = 0$ ;
- ▶ an optimization problem which can be easily solved.

## Drawbacks

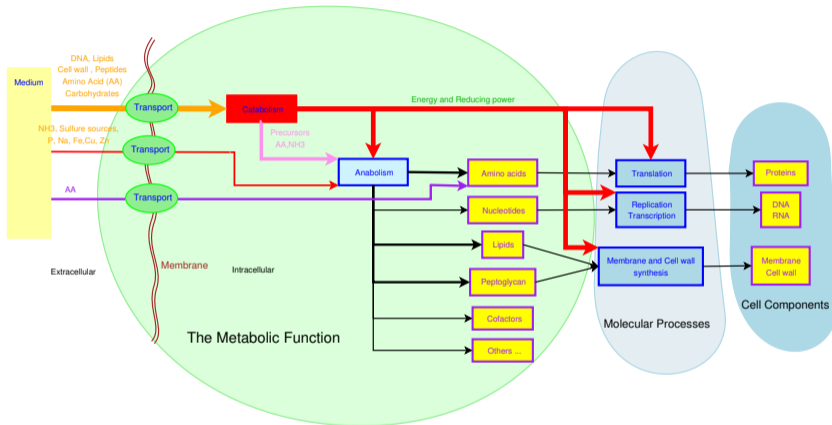
- ▶ choice of the objective function (maximization of biomass);
- ▶ need to limit superior bounds on uptake fluxes to have a solution;
- ▶ genetic regulations are not included.

**Conclusion:** FBA problem is not enough constrained.

## Include other structural constraints active during exponential growth (as mass conservation)

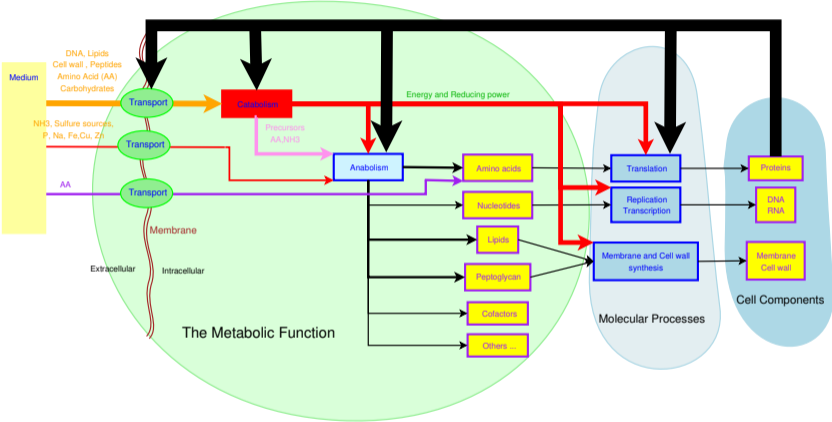
- ▶ FBAwMC for *FBA with Molecular Crowding*: Vazquez et al. *Impact of the solvent constraint on Escherichia coli metabolism*. BMC Systems Biology, 2:7. 2008.
- ▶ RBA for *Resource Balance Analysis*: Goelzer et al. *Cell design in bacteria as a convex optimization problem*. Automatica. 47(6):1210-1218. 2011.

# Description of biomass



Different molecular processes produce cellular components. . .

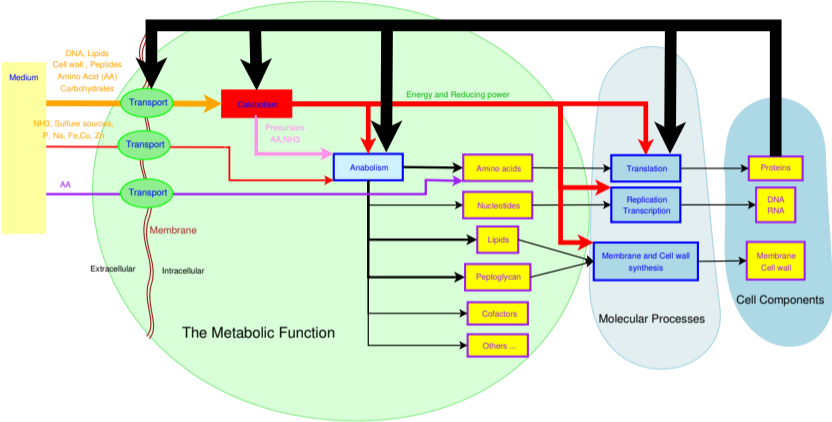
# An implicit coupling: proteins



... by sharing a common resource: proteins.



# An implicit coupling: proteins



Need to integrate the protein concentration in the optimization problem.

## Three structural constraints during exponential growth

# Exponential growth

By definition of the “**balanced**” regimen of exponential growth, we have

- ▶ the growth rate  $\mu$  is constant;

$$\dot{V}(t) = \mu V(t)$$

- ▶ the concentrations of the different cellular components are also constant.

A first consequence: for each protein  $\mathcal{P}_j$  of concentration  $P_j$ ,

$$\frac{dP_j(t)}{dt} = \underbrace{p_j(t)}_{\text{Production}} - \underbrace{\mu P_j(t)}_{\text{Dilution}}$$

$\implies$  a constant flux of protein production of  $p_j = \mu P_j \neq 0$  is required in exponential growth (steady-state).

For all proteins, a total flux equal to  $\sum_j p_j = \mu \sum_j P_j$  of protein production is required.

# A first structural constraint

Ribosomes are composed of proteins.

Consequence 2: for the ribosomes of concentration  $R$ ,

- ▶ a synthesis flux of  $r = \mu R$  is required in steady-state;
- ▶ which leads to a constraint on the capability of the protein production

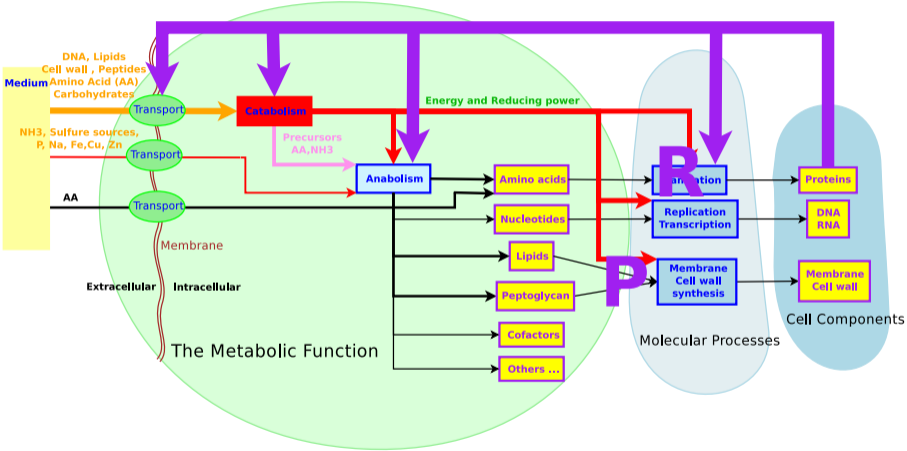
$$\mu R \leq k_T R \implies \mu \leq k_T.$$

$k_T$ : is related to the efficiency of the translation of ribosomes.

A first structural constraint on the capability of protein production by the translation apparatus

Let us now distinguish another set of proteins . . .

# Another set of proteins $P$



# Impact of nonribosomal proteins

Let us consider another set of proteins  $\mathcal{P}$  of concentration  $P$ .

$k_T$ , an optimistic superior bound ?

$$\mu(R + P) \leq k_T R \quad \Longrightarrow \quad \mu \leq k_T \frac{R}{R + P}$$

$\Longrightarrow$ the superior bound is decreasing.

# Impact of variations of $P$

**Objective:** Mimic the repression of the protein synthesis.

- ▶ Initial state of given concentration  $P_1$

$$\mu(R + P_1) \leq k_T R$$

- ▶ Let  $\delta P \in [0, P_1)$  such as  $P_2 = P_1 - \delta P$ .

$$\mu(R + P_2 + \delta P) \leq k_T R$$

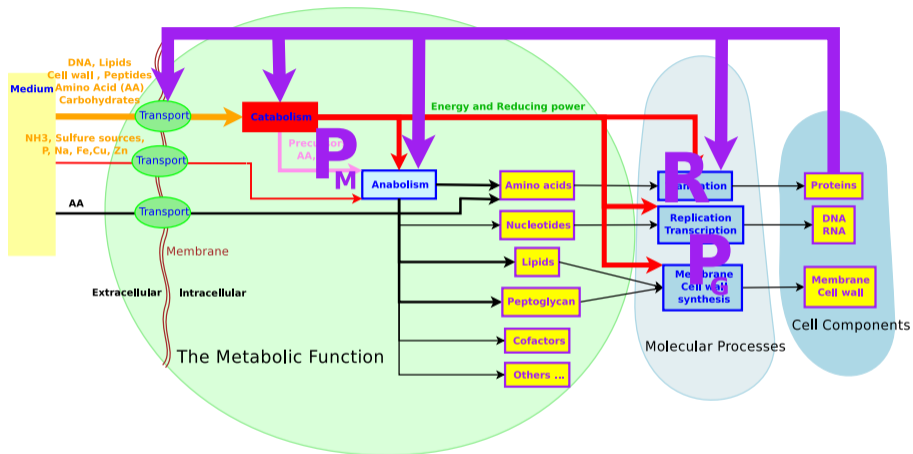
- ▶ by choosing  $\delta\mu = \mu \frac{\delta P}{R + P_2}$ ,

$$(\mu + \delta\mu)(R + P_2) \leq k_T R$$

$\implies \mu$  can increase.



# Two sets of nonribosomal proteins



- ▶  $\mathcal{P}_M$  involved in the metabolic network;
- ▶  $\mathcal{P}_G$  belonging neither to  $\mathcal{P}_M$  nor to  $\mathcal{R}$ .

## A second structural constraint

Let  $\nu$  the flux of metabolic precursors produced by  $\mathcal{P}_M$ . So

$$\nu = \mu(\alpha_R R + \alpha_G P_G + \alpha_M P_M)$$

where

- ▶  $\alpha_R$  is the amount of precursors required to ribosome synthesis;
- ▶  $\alpha_M$  is the amount of precursors required to  $\mathcal{P}_M$  synthesis;
- ▶  $\alpha_G$  is the amount of precursors required to  $\mathcal{P}_G$  synthesis.

A second structural constraint on the capability of production of the metabolic network to provide the metabolic precursors required for the cell growth

# An important assumption

Integrate the link between flux and enzyme concentration.

Recall that  $\nu = f_M(S, P)P_M$ , where  $f_M(S, P)$  is the apparent catalytic rate (also called apparent turnover rate) of the enzyme.

**Assumption:** for a given exponential growth condition, metabolite concentrations are constant. So we simplified the non-linear relation  $f_M(S, P)$  by one parameter  $k_M$ .

- ▶ A mass balance constraint.

$$\nu = \mu(\alpha_R R + \alpha_G P_G + \alpha_M P_M)$$

- ▶ A capacity constraint.

$$-k'_M P_M \leq \nu \leq k_M P_M$$

where  $k_M$  and  $k'_M$  are the apparent catalytic rate in the forward and backward direction of  $\mathcal{P}_M$ .

## A third structural constraint

Let us return to

$$\mu(R + P) \leq k_T R \quad \Longrightarrow \quad \mu \leq k_T \frac{R}{R + P}$$

Theoretically, we could have

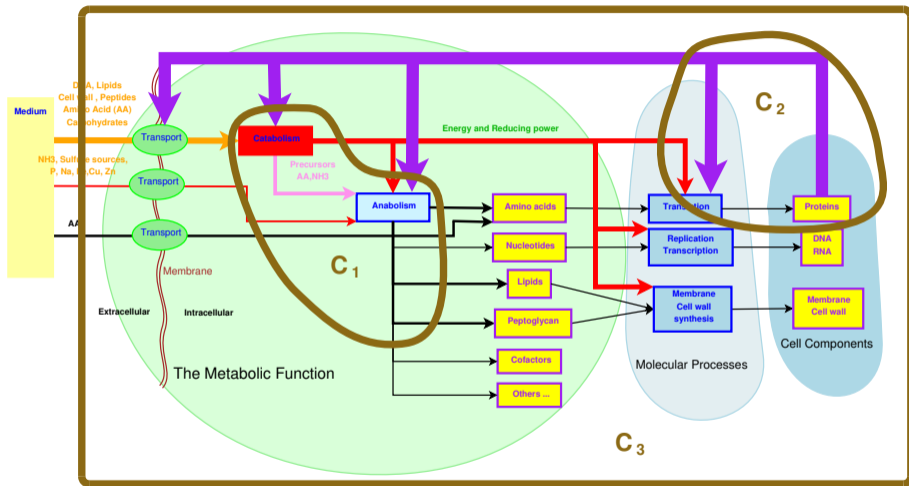
$$\lim_{R \rightarrow \infty} k_T \frac{R}{R + P} = k_T.$$

**BUT:** The intracellular volume is limited.

$\Longrightarrow$  Every cellular component has a maximal concentration.

A third structural constraint on the volume of cellular compartments and membrane occupancy

# Three structural constraints

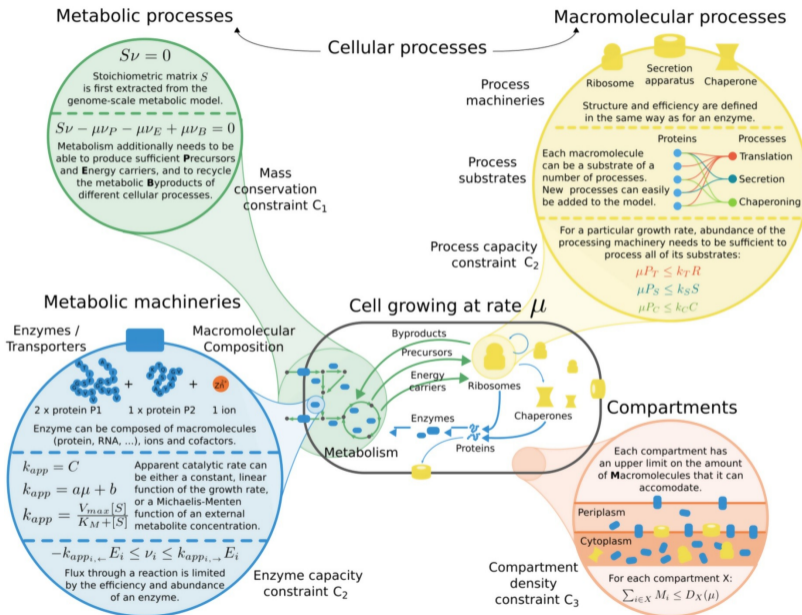


And the mass conservation and the stoichiometry of metabolic network

## Generalization to the whole cell

# Resource Balance Analysis in a nutshell

At steady state  
(constant growth rate)



# The RBA optimisation problem

Abundance of  
housekeeping proteins

Growth rate

Abundance of  
molecular machines

For fixed  $P_G \geq 0, \mu \geq 0,$

Metabolic fluxes

Fixed abundances of macro-components  
(i.e. cell wall precursors, etc.)

find  
subject to

$$Y \in \mathbb{R}_{\geq 0}^{m+p}, \nu \in \mathbb{R}^m,$$

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$$

$$(C_{2a}) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

$$(C_{2b}) \quad -K'_E Y \leq \nu \leq K_E Y$$

$$(C_3) \quad C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$$

Efficiencies of  
molecular machines  
(i.e. kappa)

Densities of compartments



# The RBA optimisation problem

Abundance of housekeeping proteins

Growth rate

Abundance of molecular machines

Metabolic fluxes

For fixed  $P_G \geq 0, \mu \geq 0,$

find

subject to

$$Y \in \mathbb{R}_{\geq 0}^{m+p}, \nu \in \mathbb{R}^m,$$

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$$

Medium setting

$$(C_{2a}) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

$$(C_{2b}) \quad -K'_E Y \leq \nu \leq K_E Y$$

(for a transporter uptaking a metabolite  $S_{ext}$ )

$$K_E = \frac{k_{max} S_{ext}}{K_m + S_{ext}}$$

Abundance of

$S_{ext}$

$$(C_3) \quad C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$$

# Theoretical properties of $P_f(\mu)$

The following theoretical properties are obtained:

1.  $P_f(\mu)$  is a Linear Programming problem;  
 $\implies$  same complexity as the FBA problem, the RBA problem is easily solvable!
2. For a set of given environmental conditions, **there exists a maximum growth rate**
  - ▶ **without defining any objective function** (by contrast to FBA);
  - ▶ **defined by a trade-off on proteins**;
  - ▶ for which a protein distribution (enzyme/ribosome) exists;
  - ▶ which can be computed by dichotomy with  $P_f(\mu)$ ;
3. Theoretical predictions of induced/repressed metabolic sub-systems with respect to the concentrations of extracellular nutrients;
4. Every mechanism allowing to save proteins increases the growth rate.

# What predicts RBA?

For a set of given extracellular concentration of nutrients, RBA computes

- ▶ the maximal growth rate
- ▶ the metabolic fluxes including the substrate uptake and by-product secretion rates
- ▶ the abundances of molecular machines such as enzymes, transporters, chaperones and proteins involved in the translation apparatus

But if an objective function is added, one can look for the cellular configuration maximizing the production of a compound (metabolite, or protein) of interest at given growth rate.

Prediction of the Monod's curve  $\mu = f(\text{Glucose})$

# RBA captures the macroscopic/microscopic behavior of bacteria

## Monod's experiments

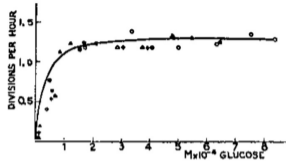
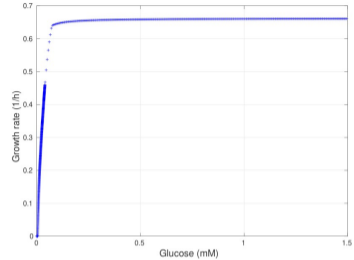


FIG. 4.—Growth rate of *E. coli* in synthetic medium as a function of glucose concentration. Solid line is drawn to equation (2) with  $R_K = 1.35$  divisions per hour, and  $C_1 = 0.22 \text{ M} \times 10^{-4}$  (11). Temperature  $37^\circ \text{ C}$ .



Prediction of the relation Ribosome =  $f(\mu)$

# RBA captures the macroscopic/microscopic behavior of bacteria

## Monod's experiments

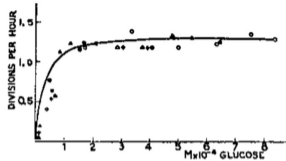
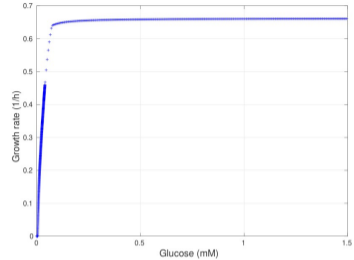
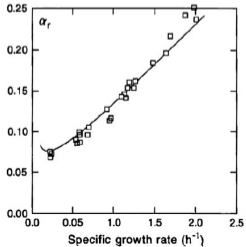


FIG. 4.—Growth rate of *E. coli* in synthetic medium as a function of glucose concentration. Solid line is drawn to equation (2) with  $R_K = 1.35$  divisions per hour, and  $C_1 = 0.22 \text{ M} \times 10^{-4}$  (11). Temperature  $37^\circ \text{ C}$ .



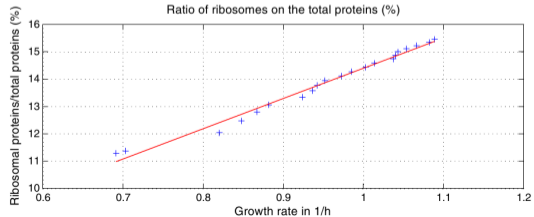
## Marr's experiments

### Ribosomes



$1 - \alpha_r$

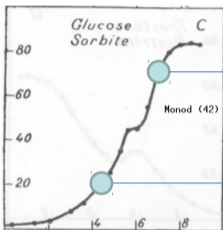
A. Goelzer, V. Fromion / *Biochimica et Biophysica Acta* 1810 (2011) 978–988



## Prediction of a hierarchy in the use of nutrients



# Monod's experiments recovered by RBA



Predicted use of carbon source by RBA

Sorbitol only

Glucose only

Predicted use of carbon source by other constraint-based modeling methods (FBA)

Sorbitol + Glucose

Sorbitol + Glucose

RBA predicts a hierarchy in the use of carbon sources

Most induced/repressed sub-systems in the metabolic network coincides with a known regulatory structure



Promising to predict the cell phenotype in combined stress conditions

## Estimation of model parameters

## Parameters to be estimated

For fixed  $P_G \geq 0$ ,  $\mu \geq 0$ ,

find  $Y \in \mathbb{R}_{\geq 0}^{m+p}$ ,  $\nu \in \mathbb{R}^m$ ,  
subject to

(C<sub>1</sub>)  $-\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$

(C<sub>2a</sub>)  $\mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$

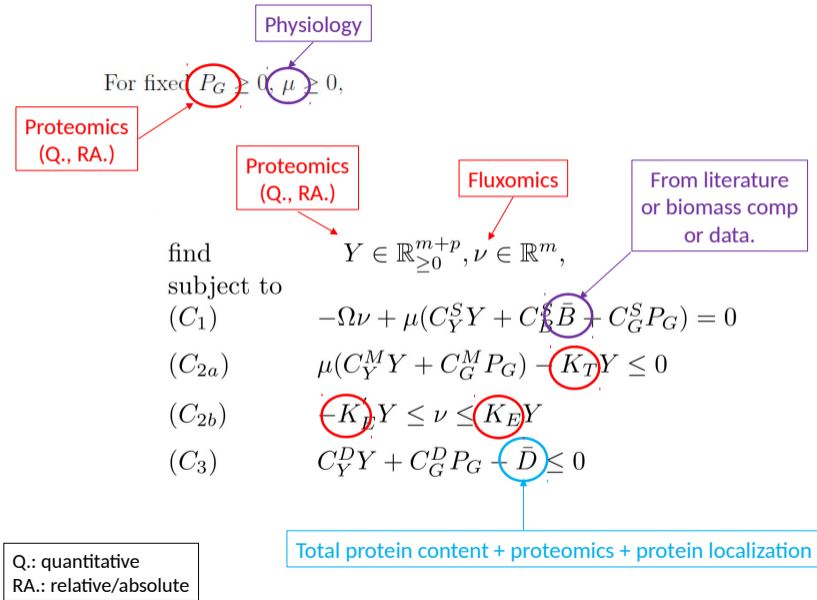
(C<sub>2b</sub>)  $-K'_E Y \leq \nu \leq K_E Y$

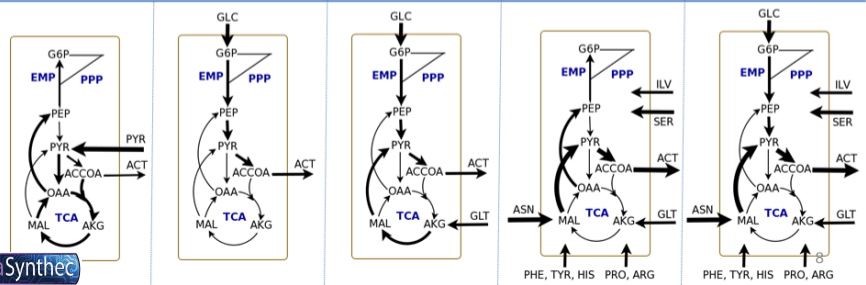
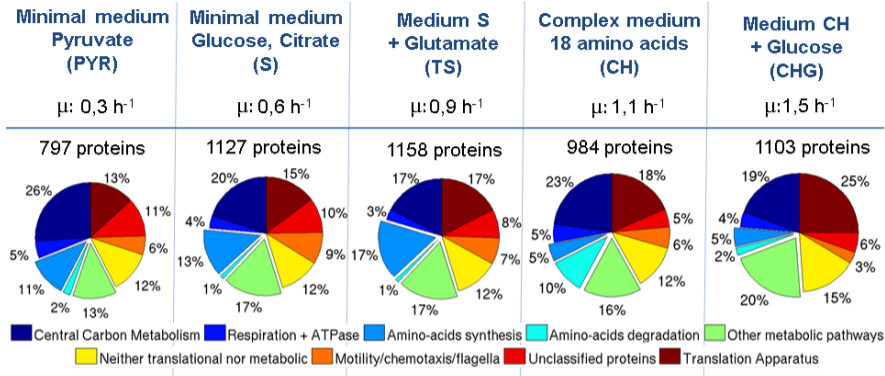
(C<sub>3</sub>)  $C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$

From the stoichiometry  
of chemical reactions

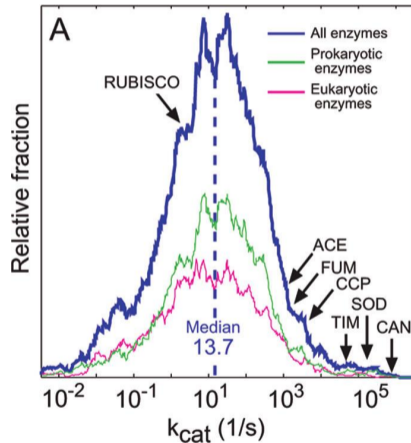
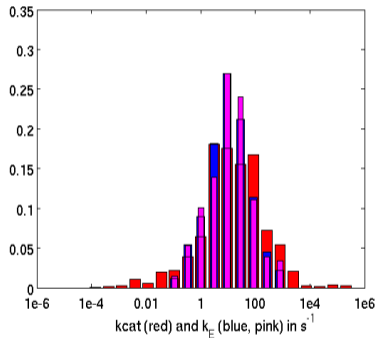
From annotation  
& bioinformatics

## Parameters to be estimated



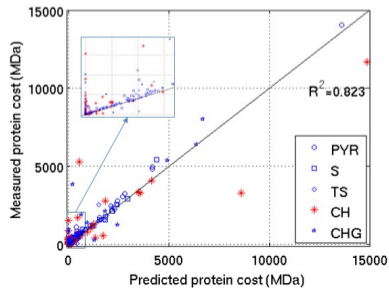
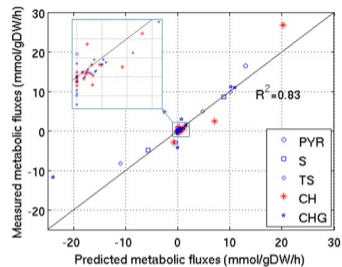
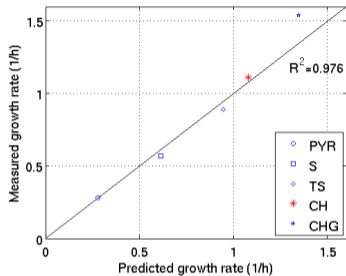


# Identification of apparent catalytic rate of 600 enzymes (Consistency with the expected distribution)



A. Bar-Even, et al. The Moderately Efficient Enzyme: Evolutionary and Physicochemical Trends Shaping Enzyme Parameters, *Biochemistry*, 2011, 50 (21), pp. 4402-4410

## Quantitative prediction of the resource allocation



## Generation of RBA models



# How to build a resource allocation model for another bacterium?

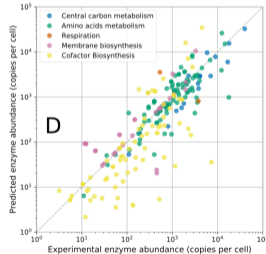
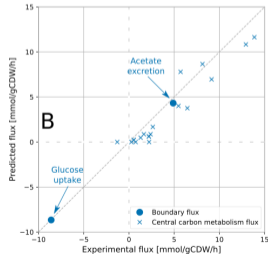
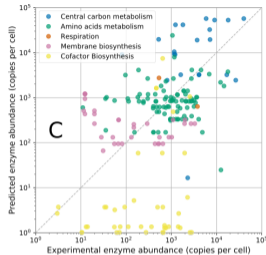
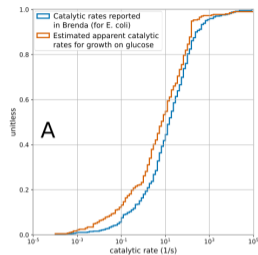
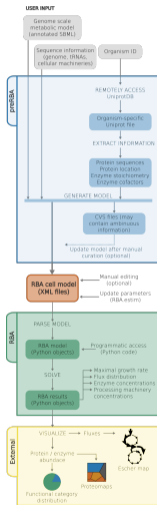
## For model building, one needs

- ▶ the annotated genomic sequence
- ▶ a genome-scale metabolic model in SBML format
- ▶ description of the molecular machines of the macro-molecular processes that you want to include in RBA

## For model calibration, one needs

- ▶ quantitative proteomic datasets
- ▶ uptake and excretion rate of nutrients, fluxomics

# Use RBApy to generate a calibrated RBA model



# Conclusion

The RBA framework provides theoretical properties for the cell design in bacteria

- ▶ The trade-off on resources structurally limits the growth rate;
- ▶ Every mechanism allowing to save resources increases the growth rate.

The RBA framework can efficiently predict for a set of concentrations of extracellular nutrients

- ▶ The maximal growth rate;
- ▶ The resource repartition of the cell: the abundance of enzymes, transporters and non-metabolic molecular machines as ribosomes and metabolic fluxes;
- ▶ The impact of a rational modification of the bacterial strain (e.g. plug-in an entire metabolic pathway).

RBAPy makes RBA model generation and simulation accessible for a large diversity of prokaryotes

see <https://rba.inrae.fr/> for existing RBA models and code availability.

We are hiring ! (30 months position to develop a RBA model of cancerous and healthy cells)